SESQUITERPENES IN LEAF POCKET RESINS OF COPAIFERA SPECIES

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Abstract—Four sesquiterpene leaf resin components were isolated and identified from Copaifera leaf resin. Additional GC and mass spectrometric evidence support the close similarity of Copaifera leaf pocket resin composition with that of the related genus, Hymenaea.

INTRODUCTION

Analysis of the sesquiterpenes in the genus Copaifera is part of a long-range investigation of the evolution of tropical leguminous resin-producing genera [1, 2]. Copaifera is an amphi-Atlantic leguminous genus and a member of the subfamily Caesalpinioideae and the tribe Detarieae. At present ca 30 species of Copaifera are recognized to occur in the New World [3, 4] and four in Africa [5]. A monographic revision is also currently in progress [Langenheim, J. H. and Freitas da Silva, M., unpublished results]. Copaifera is related to the genus Hymenaea, the resin chemistry, systematics and ecology of which have been studied in some detail in our laboratory [6-16]. In Africa, Hymenaea occurs exclusively in eastern coastal areas, whereas Copaifera is distributed only in the west; however, in the New World the two genera have essentially the same distribution with numerous species co-occurring throughout the lowland tropical ecosystems [17]. These distributional relationships, the occurrence of resin from both genera as amber (fossilized resin) and their general resin chemistry make the comparison of their evolution particularly interesting [1, 16, 17]. There are both striking similarities as well as differences in the resin chemistry and secretory structures of all organs of the two genera. Leaf resin in both genera is secreted from epithelial cells which line small, ovoid, schizogenouslyproduced pockets [18, 19]. Ten of ca 15 hydrocarbons have been identified from Hymenaea leaf resins with all species containing essentially the same compounds [6]. The leaf resin constituents are generally comprised of sesquiterpene hydrocarbons, although in some cases there may be a few oxygenated sesquiterpenoids. All of the sesquiterpenes identified in Hymenaea have also been identified in Copaifera wood resin [2, 20].

RESULTS AND DISCUSSION

Leaf resins representative of Copaifera were obtained from C. officinalis L. and C. venezuelana var. laxa Xena and Arroyo by steam distillation. Resin from C. officinalis was initially separated by prep. GC and the isolated components identified using IR. Only two compounds, γ -cadinene and caryophyllene were sufficiently purified in

this case for conclusive IR identification due to the difficulty of separating the structurally closely related resin components by prep. GC alone. Therefore, in continued work, resin isolated from Copaifera venezuelana was first separated by CC on silver nitrate impregnated silica gel; then isolated compounds were further purified by prep. GC. Two additional sesquiterpenes, cyperene and α-copaene, were thus isolated and identified by IR. Further confirmation of the identities of these four compounds was obtained by GC and GC/MS of the purified components and by analysis of resin mixtures in which they occur.

On the basis of GC retention times established during routine analysis of many different samples separated on analytical packed Carbowax columns, the constituents of Copaifera appear to be the same as those in Hymenaea leaf resins; cochromatographed samples of Hymenaea and Copaifera leaf resins on these packed columns further show that the retention times are virtually the same for all of the constituents previously identified in Hymenaea. Additionally, analytical GC on glass open-tubular columns of polar (Carbowax 20M) and apolar (SP-2100) phases and GC/MS were used to compare identified Hymenaea leaf sesquiterpenes with the remaining unisolated components of Copaifera. In this system peaks corresponding to α -cubebene, α -copaene and β -humulene occur in resins from both genera. Further indications for the presence of γ -muurolene and β -selinene were found with the open tubular Carbowax column. However, some discrepancies for these compounds occurred when compared on the SP-2100 open tubular column. y-Muurolene was not distinguishable on this column, which could be due either to lack of resolution on this apolar phase or to some alteration of γ -muurolene. Also, only α -selinene was resolved on this column. The presence of both selinene isomers—with different ones appearing on the two columns—may be explained by on-column interconversion, which has sometimes been observed on Carbowax packed columns in our previous studies of Hymenaea.

EXPERIMENTAL

Sampling. 765 g (dry wt) leaves of Copaifera officinalis collected near Mochima, El Sucre, Venezuela (collection No. JHL 6124)

and 896 g (dry wt) Copaifera venezuelana collected near Machiques, Zulia, Venezuela (collection No. JHL 6139), were powdered in a Waring blender and steam distilled using a Likens-Nickerson apparatus modified according to Maarse and Kepner [21].

Separation. The resin distillate from Copaifera was separated by prep. GC using 10% Carbowax 20M, 4 m × 5 mm, isothermal at 165°, TC. Resin from Copaifera venezuelana was initially passed through a basic alumina column eluting with n-pentane to remove oxygenated compounds. It was then separated on AgNO₃–Si gel eluting with light petrol (bp 37–55°), followed by cyclohexane-petrol mixtures (1:100–1:1) then C_6H_6 -n-pentane mixtures (1:100–1:1) and finally $Et_2O-C_6H_6$ mixtures (1:100–1:5). These fractions were finally purified using prep. GC. Collected fractions were analysed by analytical GC (2.5% Carbowax 20M on Gas Chrom Q; 8 m × 0.3 mm; 130°; N₂ 26 ml/min; FID).

Further analytical GC was carried out on WCOT columns coated with Carbowax 20M [22], $60\,\mathrm{m}\times0.3\,\mathrm{mm}$; and SP-2100; $31\,\mathrm{m}\times0.25\,\mathrm{mm}$, splitting injector temp. 165° , split ratio 1:50, column temp. 120.5° , isothermal, N₂ $12.5\,\mathrm{cm/sec}$, FID temp. 200° . GC/MS was carried out with the above columns, column temp. 135° , H₂ $20\,\mathrm{cm/sec}$, Finnigan 3200, ion source temp. 200° , $70\,\mathrm{eV}$, Series $6000\,\mathrm{data}$ system. In addition to analysis of isolated sesquiterpenes, resin samples from *C. officinalis* (collection No. JHL $6124\,\mathrm{and}$ 6126) were compared with a pooled resin sample from *H. courbaril* L. from Guerrero, Mexico (collection Nos. JHL 4969, 5014, 5015) by open tubular GC and GC/MS.

IR spectra were obtained neat using 0.1 mm microcells and a Perkin-Elmer 237-B spectrophotometer grating IR (Barnes 0.1 mm microcell with beam condenser). Detailed comparison of IR spectra with literature was used for identification [23].

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REFERENCES

- 1. Langenheim, J. H. (1969) Science 163, 1157.
- Langenheim, J. H. (1981) in Advances in Legume Systematics (Polhill, R. M. and Raven, P. H., eds.) p. 627. Royal Botanical Gardens, Kew.
- 3. Dwyer, J. D. (1951) Brittonia 7, 143.
- 4. Xena, N., Arroyo, M. T. and Langenheim, J. H. Acta Bot. Venez. (in press).
- 5. Léonard, J. (1957) Mem. Acad. R. Belg., Cl. Sci. 30, 1.
- Martin, S. S., Langenheim, J. H. and Zavarin, E. (1972) Phytochemistry 11, 3049.
- 7. Martin, S. S., Langenheim, J. H. and Zavarin, E. (1974) Biochem. Syst. Ecol. 3, 760.
- 8. Martin, S. S., Langenheim, J. H. and Zavarin, E. (1976) Biochem. Syst. Ecol. 4, 181.
- 9. Lee, Y. T. and Langenheim, J. H. (1975) Univ. Calif. Berkeley Publ. Botany 69, 1.
- Langenheim, J. H., Stubblebine, W. H., Lincoln, D. E. and Foster, C. E. (1978) Biochem. Syst. Ecol. 6, 299.
- 11. Langenheim, J. H., Stubblebine, W. H. and Foster, C. E. (1979) Biochem. Syst. Ecol. 7, 21.
- Langenheim, J. H., Arrhenius, S. P. and Nascimento, J. C. (1981) Biochem. Syst. Ecol. 9, 27.
- Langenheim, J. H., Foster, C. E. and McGinley, R. B., (1980) Biochem. Syst. Ecol. 8, 385.
- 14. Stubblebine, W. H., Langenheim, J. H. and Lincoln, D. E. (1975) Biochem. Syst. Ecol. 3, 219.
- Stubblebine, W. H. and Langenheim, J. H. (1977) J. Chem. Ecol. 3, 633.
- Langenheim, J. H. (1972) Publicações Culturais. Companhia de Diamantes de Angola 85, 15.
- Langenheim, J. H. (1973) in Tropical Forest Ecosystems in Africa and South America: A Comparative Review (Meggers B. J., Ayensu, E. S. and Duckworth, W. D., eds.) p. 89. Smithsonian Press, Washington.
- Langenheim, J. H. (1967) J. Arnold Arbor. Harv. Univ. 48, 203.
- Langenheim, J. H., Lincoln, D. E., Stubblebine, W. H. and Gabrielli, A. C. (1982) Am. J. Botany 69, 595.
- Wenninger, J. A., Yates, R. L. and Dolinsky, M. (1967) J. Assoc. Off. Analyt. Chem. 50, 1304.
- Maarse, H. and Kepner, R. E. (1970) J. Agric. Food Chem. 18, 1095.
- 22. Grob, K. and Grob, D. (1976) J. Chromatogr. 125, 471.
- Wenninger, J. A., Yates, R. L. and Dolinsky, M. (1967) J. Assoc. Off. Analyt. Chem. 50, 1313.